

Clinical and Translational Gastroenterology Publish Ahead of Print DOI: 10.14309/ctg.000000000000656

Determination of Optimal Eosinophil Thresholds for Diagnosis of Eosinophilic Gastritis and Duodenitis: A Pooled Analysis of 4 Prospective Studies

Evan S. Dellon, MD, MPH¹; Enoch Bortey, PhD²; Alan T. Chang²; Craig A. Paterson, MD²;

Kevin Turner, DO³; Robert M. Genta, MD⁴

¹University of North Carolina School of Medicine, Chapel Hill, NC; ²Allakos Inc., San Carlos, CA;

³University of Texas Southwestern Medical Center, Dallas, TX; ⁴Baylor College of Medicine,

Houston, TX

Journal: Am J Gastroenterol

Suggested running head: Optimal eosinophil thresholds

Word count (3,000 words, not including references, tables, or abstract): 2704

Keywords: Eosinophilic gastrointestinal diseases, eosinophil count, high-power field, stomach,

duodenum

This is an open access article distributed under the terms of the Creative Commons Attribution-NonCommercial-NoDerivatives License 4.0 (CC BY-NC-ND), which permits downloading and sharing the work provided it is properly cited. The work cannot be changed in any way or used commercially without permission from the journal. Guarantor of the article: Evan S. Dellon, MD, MPH

Specific author contributions: ESD contributed to the planning and conduct of the study, collection and interpretation of data, drafting of the manuscript, and approval of the final draft. EB contributed to the planning and conduct of the study, collection and interpretation of data, drafting of the manuscript, and approval of the final draft. ATC contributed to the planning and conduct of the study, collection and interpretation of data, drafting of the study, collection and interpretation of data, drafting of the study, collection and interpretation of data, drafting of the final draft. CAP contributed to the planning and conduct of the study, collection and interpretation of data, and approval of the final draft. KT contributed to the collection and interpretation of data, drafting of the manuscript, and approval of the final draft. RMG contributed to the collection and interpretation of data, drafting of the manuscript, and approval of the final draft.

Financial support: This study was supported by Allakos, Inc.

Potential competing interests: ESD has received research funding from Adare/Ellodi, Allakos, Arena, AstraZeneca, Celgene/Receptos/BMS, GSK, Meritage, Miraca, Nutricia, Regeneron, Revolo, and Shire/Takeda; consults for Abbott, AbbVie, Adare/Ellodi, Aimmune, Akesobio, ALK, Allakos, Amgen, Arena, Aslan, AstraZeneca, Avir, Biorasi, Calypso, Celgene/Receptos/BMS, Celldex, Eli Lilly, EsoCap, Eupraxia, GSK, Gossamer Bio, Invea, Landos, LucidDx, Morphic, Nutricia, Parexel/Calyx, Phathom, Regeneron, Revolo, Robarts/Alimentiv, Salix, Sanofi, Shire/Takeda, and Target RWE; and has received educational grants from Allakos, Banner, and Holoclara. EB, ATC, and CAP are employees of and may own stock or stock options in Allakos. KT has consulted for Adare and Allakos. RMG has consulted for Adare/Ellodi, Allakos, NexEos Diagnostics, Red Hill Pharmaceuticals, Revolo, and Sanofi.

Correspondence and reprint requests to:

Evan S. Dellon, MD, MPH CB#7080 Bioinformatics Bldg. 130 Mason Farm Rd. Chapel Hill, NC 27599-7080 Email: evan_dellon@med.unc.edu

ABSTRACT

Objectives: Consensus is lacking regarding the number of eosinophils (eos) required for diagnosis of eosinophilic gastritis (EoG) and eosinophilic duodenitis (EoD). Additionally, thresholds that require multiple high-power fields (hpfs) may not be practical for clinical use, resulting in delayed or missed diagnoses. This pooled analysis of 4 prospective studies assessed thresholds for multiple and single hpfs used to diagnose EoG and EoD. **Methods:** Studies included the phase 2 ENIGMA1, the phase 3 ENIGMA2, an EoG/EoD prevalence study, and a healthy volunteer study. Eosinophils were quantified in the epithelium and lamina propria for controls and symptomatic participants. Symptomatic participants were further divided by histologic diagnosis of EoG/EoD. Peak eos counts were assessed, and area under the receiver operating characteristic (AUROC) curve was analyzed to identify eos cutoffs for detection of EoG/EoD using the Youden index and sensitivity and specificity equality approaches.

Results: Based on the highest specificity analysis in 740 patients, the optimal eos threshold was determined to be 20 eos/hpf in 5 gastric hpfs for EoG (71% sensitivity; 94% specificity) and 33 eos/hpf in 3 duodenal hpfs for EoD (49% sensitivity; 100% specificity). For single-field analysis, the optimal eos thresholds were 33 eos/hpf (EoG) and 37 eos/hpf (EoD), both corresponding to 93% sensitivity and 93% specificity.

Conclusions: Highly specific single gastric and duodenal hpf thresholds may have more clinical applicability than thresholds requiring multiple hpfs and could better facilitate development of practical histopathologic guidelines to aid pathologists and clinicians in the detection and diagnosis of EoG and/or EoD.

Word count (limit 250): 248

STUDY HIGHLIGHTS

What is known

- Diagnosis of eosinophilic gastritis and duodenitis is challenging because guidelines regarding eosinophil count thresholds are lacking
- Assessment of multiple high-power fields when quantifying eosinophils is timeconsuming and may be clinically impractical

What is new here

- Pooled-data analysis identified highly specific eosinophil count thresholds for diagnosing eosinophilic gastritis and duodenitis with a single high-power field
- These single high-power field thresholds may have more clinical applicability than thresholds requiring multiple high-power fields
- These thresholds can be used to inform future diagnostic guidelines

INTRODUCTION

Eosinophilic gastrointestinal (GI) diseases (EGIDs) are characterized by GI symptoms and the pathologic infiltration of eosinophils (eos) in the absence of another identified cause (1-3). The term EGID encompasses several specific eosinophilic disorders, including eosinophilic esophagitis (EoE), eosinophilic gastritis (EoG), and eosinophilic duodenitis (EoD) (1). Diagnosis of EGIDs can be challenging because symptoms (e.g., nausea, abdominal pain, vomiting, early satiety, diarrhea, weight loss, food allergy/intolerance) are nonspecific and often overlap with other GI conditions (4, 5). In addition, diagnosis is often delayed owing to an absence of diagnostic guidelines for non-EoE EGIDs in adult patients (1, 5-7). There is also a lack of disease awareness related to low suspicion of EGIDs during workup, insufficient number and location of biopsies obtained from esophagogastroduodenoscopies (EGD), failure to obtain biopsies during EGD, and failure to enumerate eos in gastric and duodenal biopsies (5, 8-10).

Further complicating the diagnostic process is the lack of consensus among experts regarding the number of eos required for diagnosis. Several studies have used varying eos count thresholds suggested by expert opinion and literature review, including for stomach (e.g., \geq 30 eos in 5 high-power fields [hpfs] (11), \geq 30 eos/1 hpf (4), \geq 70 eos/hpf (12), \geq 20 eos/hpf (13, 14)) and duodenum/small bowel (e.g., \geq 52 eos/hpf (11), \geq 30-50 eos/hpf (4), \geq 30 eos/hpf (14), \geq 20 eos/hpf (13, 15)). However, studies examining eos thresholds are somewhat limited, as they rely on retrospective approaches or samples from patients who were undergoing endoscopy for a clinical indication but had "normal" samples, which might lead to biased results (16). The few studies of autopsy data may be unreliable because rapidly occurring postmortem autolytic changes preclude an accurate histologic assessment of the gastrointestinal mucosa (17, 18).

Although for clinical trials, the US Food and Drug Administration has accepted cutoffs of \geq 30 eos/hpf in \geq 5 hpfs for the diagnosis of EoG and in \geq 3 hpfs for the diagnosis of EoD, the threshold requirements for regulatory approval of therapeutics may not necessarily reflect

thresholds used in clinical practice; furthermore, time constraints of the histopathologic examination make thresholds requiring counting eos in multiple hpfs impractical (5). Accordingly, an evidence-based approach is needed to establish appropriate eos thresholds for the diagnosis of EoG and EoD to improve the detection of patients with EGIDs and decrease the time to diagnosis. Therefore, the primary objectives of this study were to assess thresholds that have used multiple hpfs to diagnose EoG and EoD, compare them to thresholds used in clinical trials, and determine reliable thresholds for a single hpf.

METHODS

Participant Population

This was a pooled analysis of 4 prospective studies that included 3 clinical trials of adult participants (≥18 years old) with moderate-to-severe, chronic unexplained GI symptoms (phase 2 ENIGMA1 [NCT03496571] (19), phase 3 ENIGMA2 [NCT04322604] (20), and a study to identify the prevalence of EoG and EoD) (21) and a healthy volunteer study (21). All studies were performed in accordance with the Declaration of Helsinki and in compliance with Good Clinical Practice Guidelines, the institutional review boards of the trial sites, and applicable laws. All participants provided written informed consent before trial entry.

Symptom entry criteria from the phase 3 ENIGMA2 study (20) were applied to identify participants for this pooled analysis. Patients with the following symptoms were screened: bloating, nausea, early satiety, loss of appetite, and abdominal cramping and abdominal pain. After we excluded individuals with *Helicobacter pylori* and/or celiac disease, participants from the ENIGMA1, ENIGMA2, and prevalence studies were eligible if they had weekly average total symptom scores (TSS6; score range: 0-60) ≥10 for at least 2 weeks of screening and abdominal pain, diarrhea, and/or nausea scores ≥3 at baseline. The TSS6 was calculated from a daily patient-reported outcome questionnaire on which participants rate the presence and severity (on

a scale of 1 to 10) of bloating, nausea, early satiety, loss of appetite, and abdominal cramping and pain; this scale was similar to that utilized in prior trials (TSS8, which included vomiting and diarrhea, not reported in the TSS6) (19). These symptomatic participants who exhibited TSS of ≥10 underwent an EGD with biopsy (Supplementary Table). Controls from the healthy volunteer study had TSS6 scores ≤1, indicating no active GI symptoms when they underwent EGD with biopsy.

Eosinophil Quantification

The systematic eos quantification methodology that we used has been described previously (5). Multiple biopsies (≥12) were taken from all participants according to a standardized protocol: 8-10 gastric biopsies (including at least 4 from the antrum and 4 from the body), 4-6 duodenal biopsies, and up to 4 esophageal biopsies (only if the patient had a history of EoE or if EoE features were observed during EGD) (19, 21, 22). For the purpose of this study, biopsy results from the initial screening endoscopy (for the trials) or from the single endoscopy (for the prevalence study) were evaluated for all participants.

In brief, biopsy sections were first surveyed at low power (40X and 100X) to evaluate how much tissue was present and note any unexpected pathology. Sections were then viewed at medium power (200X), and the most eosinophil-dense areas in each fragment were identified; in these areas, chosen whenever possible among the best-oriented portions of the mucosa, 5 nonoverlapping hpfs (400X, each measuring 0.237 mm²) were selected for counting. Eosinophils were counted only in the mucosa (i.e., epithelium and lamina propria). Eosinophils in the *muscularis mucosae*, submucosa, or *muscularis propria* were not counted.

Participant Stratification

On the basis of TSS scores and eos counts, participants were stratified into a "control" group, which comprised asymptomatic, healthy volunteers (n=33) with a TSS6 score <1, and a "patient" group, which comprised symptomatic participants and included a pooled population from the ENIGMA1, ENIGMA2, and prevalence study (n=707). These symptomatic participants were then further divided by histologic diagnosis. Participants met histologic criteria for EoG if they had \geq 30 eos/hpf in \geq 5 gastric hpfs and met histologic criteria for EoD if they had \geq 30 eos/hpf in \geq 5 gastric hpfs and met histologic criteria for EoD if they had \geq 30 eos/hpf in \geq 5 gastric hpfs and met histologic criteria for EoD if diagnostic criteria.

Statistical Analysis

The distribution of peak eos counts was assessed for each of the participant groups of interest (controls, symptomatic participants who met histologic criteria, and symptomatic participants who did not meet histologic criteria), and the mean of the peak counts was calculated. A Welch's *t* test was used to analyze differences in mean eos count and peak eos count between groups.

We analyzed the area under the receiver operating characteristic (AUROC) curve for all participants, taking into account participants' symptoms and histology, to identify the best eos cutoff while considering the sensitivity and specificity for detection of EoG and EoD. We used two approaches. First, we determined the Youden index, which indicates the point on the receiver operating characteristic (ROC) curve where sensitivity and specificity are optimized (23). The Youden index was calculated as the maximum (sensitivity + specificity – 1). A Youden index value of 1 indicates no false negatives or false positives (and is similar conceptually to an area under the curve of 1.0). Second, we used sensitivity and specificity equality calculated as the absolute value of (sensitivity – specificity), which measures the absolute difference between

sensitivity (rate of true positives) and specificity (rate of true negatives). Sensitivity and specificity are equal when the resulting calculation is 0. For both approaches, the eos peak count was the exposure, and the outcome was histologic EoD or EoG. Optimal eos thresholds were determined from the higher specificity outcome of either the Youden index or sensitivity and specificity equality analyses.

RESULTS

Participant Characteristics

In this analysis, 740 participants were included (ENIGMA1 [n=74], ENIGMA2 [n=324], EoG/EoD prevalence study [n=309], healthy control study [n=33]). Of the 707 symptomatic participants from ENIGMA1, ENIGMA2, and the prevalence study, 388 (55%) had histologic evidence of EoG and/or EoD. Upon further breakdown by disease, 53 (14%) had EoG, 231(60%) had EoD (2 were excluded for missing duodenal biopsies), and 106 had EoG and EoD (27%). Conversely, 319 (45%) did not meet histologic criteria (Supplementary Figure). Of the 33 controls, 2 (6%) had histologic evidence of EoD (21).

Participant characteristics are reported in Table 1; of note, the healthy volunteers were younger and had a lower proportion of females than the other groups. As expected by study entry criteria, the healthy volunteers (n=33) had TSS of approximately 0, compared with elevated scores in symptomatic participants (n=707; mean±standard deviation [SD], 29±10), EoG/EoD (n=388; mean±SD, 30±10), and eos <30/hpf groups (n=319; mean±SD, 29±10). Mean tissue eos counts for stomach and duodenum are presented in Figures 1 and 2, respectively. The mean (SD) gastric counts were 7 (5), 25 (34), 68 (50), and 13 (8) in the controls, symptomatic participants overall, symptomatic participants with histologic EoG, and symptomatic participants without EoG, respectively. The mean (SD) duodenal counts were 19 (7), 33 (21), 49 (19), and 19 (7) in the controls, symptomatic participants overall, symptomatic participants with histologic EoD, and symptomatic participants without EoD, respectively. Of

note, counts did not differ on the basis of concomitant proton pump inhibitor (PPI) use (data for prevalence study only; data not shown).

ROC Curve Analysis in Symptomatic Participants for 5 Gastric hpfs and 3 Duodenal hpfs

The ROC curve analysis included the 707 participants with symptoms (TSS score ≥ 10) from ENIGMA1, ENIGMA2, and the prevalence study and the 33 healthy controls. The AUROC curve for disease classification was 0.74 with 3 duodenal hpfs and 0.78 with 5 gastric hpfs (Figure 3). In the analysis for 5 gastric hpfs, the Youden index was highest (0.645) at 20 eos/5 hpfs, corresponding to a sensitivity of 71% and specificity of 94%. Equality of sensitivity and specificity (difference value, 0.002) was achieved at a threshold of 18, corresponding to a sensitivity of 76% and specificity of 76%. In the analysis for 3 duodenal hpfs, the Youden index was highest (0.487) at 33/3 hpfs, corresponding to a sensitivity of 49% and a specificity of 100%. Equality of sensitivity and specificity (difference value, 0.005) was achieved at a threshold of 24, corresponding to a sensitivity of 69% and a specificity of 69%. Based on the higher specificity analysis, the optimal eos threshold was 20 eos/hpf in 5 gastric hpfs for EoG and 33 eos/hpf in 3 duodenal hpfs for EoD.

ROC Curve Analysis in Symptomatic Participants for 1 Gastric hpf and 1 Duodenal hpf

The AUROC curve for disease classification was 0.94 with both 1 duodenal hpf and 1 gastric hpf (Figure 4). In the analysis to determine thresholds for 1 gastric hpf, the Youden index was highest (0.889) at 30 eos/hpf, corresponding to a sensitivity of 98% and specificity of 91%. Equality of sensitivity and specificity (difference value, 0.002) was achieved at a threshold of 33, corresponding to a sensitivity of 93% and specificity of 93%. In the analysis to determine thresholds for 1 duodenal hpf, the Youden index was highest (0.871) at 37 eos/hpf, corresponding to a sensitivity of 95% and specificity of 93%. Equality of sensitivity and specificity (difference value, 0.001) was achieved at a threshold of 37, corresponding to a

sensitivity of 93% and specificity of 93%. Based on the higher specificity analyses, the optimal eos threshold was 33 eos/hpf in 1 gastric hpf for EoG and 37 eos/hpf in 1 duodenal hpf for EoD.

ROC Curve Analysis for Empirically Selected Thresholds in the Duodenum

Owing to the difference in our calculated optimal duodenal threshold in 1 hpf (37 eos/hpf) from thresholds reported previously (≥50-52 eos/hpf) (4, 11), we examined the operating characteristics in 4 other potential clinical cutoff points in the duodenum. For 40 eos/hpf, the Youden index was 0.784, with a sensitivity of 82% and specificity of 96%. For 45 eos/hpf, the Youden index was 0.659, with a sensitivity of 66% and specificity of 99.8%. For 50 eos/hpf, the values were 0.500, 50%, and 100%, respectively, for the Youden index, sensitivity, and specificity, and for 52 eos/hpf, the values were 0.442, 44%, and 100%, respectively.

DISCUSSION

In combination with patient history and symptoms, quantification of eos from gastric and duodenal biopsies is required for diagnosing EoG and EoD. However, multiple different eos count thresholds have been reported in the literature, and currently there are no diagnostic guidelines in adults or evidence-based determinations of the optimal eos thresholds to support the diagnosis (1, 4-7, 11, 12, 15, 24). Lack of agreed-upon thresholds also may contribute to the delay in diagnosis, together with delayed referral to gastroenterologists, delays in obtaining EGDs, lack of thorough diagnostic evaluation, insufficient or no biopsies collected for eos quantification, and time constraints in clinical practice (5, 7, 9).

Accordingly, in this analysis, we sought to determine optimal eos thresholds in multiple and single hpfs from the stomach and duodenum for the detection of EoG and EoD by using data from 3 studies of symptomatic participants and 1 healthy volunteer study. Of symptomatic patients in these studies, only 55% met histologic criteria for EoG and/or EoD, underscoring the need for more specific histologic criteria. Our analysis based on the highest specificity revealed that thresholds for EoG of 20 eos/5 hpfs or 33 eos/1 hpf in the stomach, and thresholds for EoD of 33 eos/3 hpfs or 37 eos/1 hpf in the duodenum, minimize false-positive rates. Although the specificity analyses performed here suggest that these thresholds are valid, additional consideration should be given to which specific values align with practice considerations and implementation. For example, a single hpf duodenal count of 50 eos/hpf will have 100% specificity (i.e., no falsely categorized patients, as no controls have a duodenal count this high) at the expense of a lower sensitivity (i.e., versus our counts of 40 and 45 eos/hpf) and missing "true" patients.

Although various thresholds have been proposed, there is still little agreement on the number of eos per hpf regarded as pathologic in either gastric or duodenal mucosa (4, 11, 15, 16, 24). Part of the difficulty is that eos are normal tissue-resident cells in both the stomach and the duodenum, unlike the esophagus, where they are not normally present (11). One recommended approach to this problem is to select a value that is approximately twice that of the highest reported normal value, which led to the recommendation of 52 eos/hpf as a threshold for the duodenum (11). The suggestion for using multiple hpfs was an attempt to require more diffuse involvement and avoid an EGID diagnosis when only one focal area was involved (7). Inherent to this methodology, however, is a dependence on what normal values are, and from whom "normal" tissue is obtained. In an autopsy-based study and a populationbased study of normal volunteers undergoing endoscopy, the peak values ranged from ~5 to 12 eos/hpf for the stomach and ~16 to 19 in the duodenum (25, 26). Other studies used endoscopy controls as "normal," selecting patients who might be symptomatic but who have a normal endoscopic appearance, normal biopsy results, and no other identified GI conditions (15, 16, 24, 27-33). When these studies are summarized, peak gastric counts are generally below 10, but they can range as high as 15 to 16 eos/hpf, and peak duodenal counts range up to ~30 eos/hpf, though most reported values are lower (16). These values generally match what we found in our healthy control cohort, lending validity to our findings. One prior study assessed the diagnostic

specificity of various eos count thresholds depending on the number of hpfs required (16). It showed 98% specificity for >20 gastric eos in 1 hpf, as well as 95% specificity for >40 duodenal eos in 1 hpf. Although these results were obtained from participants in whom no abnormalities were detected by endoscopy, the values closely correspond to values reported in our healthy volunteer population. These data lend credence to our findings and potentially support more practical peak eos count thresholds in just 1 hpf. The exact threshold chosen would necessitate a balance of sensitivity, specificity, and clinical applicability. Lastly, although other conditions such as functional dyspepsia are also associated with increased duodenal eos counts, the thresholds reported in this analysis would likely exclude patients with functional dyspepsia, for whom counts have been reported to be 9 to 12 eos/hpf (26, 34, 35).

Limitations of this study include pooling of different studies rather than using one unified protocol; however, this pooled approach provided a large sample size of symptomatic individuals, and we harmonized the entry criteria for the current analysis. Additionally, while this analysis included a relatively small sample size of healthy volunteers, this group was highly characterized by detailed histologic assessments, TSS scores, and documented symptoms; was prospectively enrolled; and-to our knowledge-represents a unique resource in the field. These are asymptomatic and healthy subjects recruited from several sites who underwent purely research endoscopies expressly to obtain sufficient gastric and duodenal tissue for comparative analyses. This analysis included the use of large prospective and high-quality studies, which together produced the largest study assessing diagnostic thresholds for eos counts to date. However, given the limited clinical data available, we were unable to adequately examine the role of other important patient characteristics in the current analysis (e.g., allergic disease, other biomarkers). Although baseline PPI use was reported in 49% (32/65) of patients in ENIGMA (19) and 29% (116/405) of patients in the prevalence study (21), complete PPI data were not available for all studies. Further, although prior infection was not assessed, patients were screened and excluded if they had evidence of active Helicobacter pylori infection or celiac disease. Future research efforts are underway to better characterize clinicopathologic factors that may affect gastric and duodenal eosinophilia (e.g., PPI use) (36), though a prior study suggested that PPI use did not affect gastric or duodenal eos counts in a clinically meaningful way (16). Additionally, eos counts were determined by experts in the field using quantification methods that are published and standardized. Lastly, our analyses included multiple statistical approaches to assess thresholds, which permitted the identification of ideal thresholds and will allow future recommendations for practical thresholds.

In conclusion, in this pooled analysis of 4 prospective studies (2 clinical trials, an EoG/EoD prevalence study, and a healthy volunteer study), we developed the largest dataset yet reported to assess eos count thresholds for the diagnosis of EoG and/or EoD. Using healthy volunteers as controls, we found highly specific thresholds both for a requirement of multiple hpfs and for use of a single hpf, which may have more clinical applicability and gain broader acceptance by pathologists, thus aiding in the detection of more patients with EoG and/or EoD (8). Our data suggest that 30 eos in a single hpf in the stomach and 37 eos/hpf in the duodenum could be reasonably selected to histologically define patients with EoG/EoD. Additionally, thresholds with 100% specificity (e.g., 50 eos/hpf) may align well with implementation in clinical practice, identifying "true" patients at the cost of lower sensitivity. The thresholds identified here could additionally be used to help develop future practical and histopathologic diagnostic guidelines.

Supplemental Figure - <u>http://links.lww.com/CTG/B33</u> Supplemental Table - <u>http://links.lww.com/CTG/B34</u>

REFERENCES

1. Dellon ES, Gonsalves N, Abonia JP, et al. International consensus recommendations for eosinophilic gastrointestinal disease nomenclature. Clin Gastroenterol Hepatol 2022.

2. Gonsalves N. Eosinophilic Gastrointestinal Disorders. Clin Rev Allergy Immunol 2019;57:272-285.

3. Walker MM, Potter M, Talley NJ. Eosinophilic gastroenteritis and other eosinophilic gut diseases distal to the oesophagus. Lancet Gastroenterol Hepatol 2018;3:271-280.

4. Dellon ES. Eosinophilic Gastrointestinal Diseases Beyond Eosinophilic Esophagitis. Am J Gastroenterol 2022;117:697-700.

5. Turner KO, Collins MH, Walker MM, et al. Quantification of Mucosal Eosinophils for the Histopathologic Diagnosis of Eosinophilic Gastritis and Duodenitis: A Primer for Practicing Pathologists. Am J Surg Pathol 2022;46:557-566.

6. Chehade M, Tan J, Gehman LT. Gastroenterology Practice Patterns Contribute to Missed Diagnoses of Eosinophilic Gastritis and Duodenitis. Gastro Hep Advances 2022.

7. Chehade M, Kamboj AP, Atkins D, et al. Diagnostic Delay in Patients with Eosinophilic Gastritis and/or Duodenitis: A Population-Based Study. J Allergy Clin Immunol Pract 2021;9:2050-2059 e20.

8. Saad AJ, Genta RM, Turner KO, et al. Do General Pathologists Assess Gastric and Duodenal Eosinophilia? Arch Pathol Lab Med 2022.

9. Rothenberg ME, Hottinger SK, Gonsalves N, et al. Impressions and aspirations from the FDA GREAT VI Workshop on eosinophilic gastrointestinal disorders beyond eosinophilic esophagitis and perspectives for progress in the field. J Allergy Clin Immunol 2022;149:844-853.

10. Genta RM, Dellon ES, Turner KO. Non-oesophageal eosinophilic gastrointestinal diseases are undersuspected clinically and underdiagnosed pathologically. Alimentary Pharmacology & Therapeutics 2022.

11. Collins MH. Histopathologic features of eosinophilic esophagitis and eosinophilic gastrointestinal diseases. Gastroenterol Clin North Am 2014;43:257-268.

12. Ko HM, Morotti RA, Yershov O, et al. Eosinophilic gastritis in children: clinicopathological correlation, disease course, and response to therapy. Am J Gastroenterol 2014;109:1277-85.

13. Reed C, Woosley JT, Dellon ES. Clinical characteristics, treatment outcomes, and resource utilization in children and adults with eosinophilic gastroenteritis. Dig Liver Dis 2015;47:197-201.

14. Reed CC, Ketchem CJ, Miller TL, et al. Psychiatric Comorbidities Are Highly Prevalent in Nonesophageal Eosinophilic Gastrointestinal Diseases. Clin Gastroenterol Hepatol 2022;20:e664-e670.

15. Genta RM, Sonnenberg A, Turner K. Quantification of the duodenal eosinophil content in adults: a necessary step for an evidence-based diagnosis of duodenal eosinophilia. Aliment Pharmacol Ther 2018;47:1143-1150.

16. Reed CC, Genta RM, Youngblood BA, et al. Mast Cell and Eosinophil Counts in Gastric and Duodenal Biopsy Specimens From Patients With and Without Eosinophilic Gastroenteritis. Clin Gastroenterol Hepatol 2021;19:2102-2111.

17. Carlson JA, Middleton PJ, Szymanski MT, et al. Fatal rotavirus gastroenteritis: an analysis of 21 cases. Am J Dis Child 1978;132:477-9.

18. Thorpe E, Thomlinson JR. Autolysis and post-mortem bacteriological changes in the alimentary tract of the pig. J Pathol Bacteriol 1967;93:601-10.

19. Dellon ES, Peterson KA, Murray JA, et al. Anti-Siglec-8 Antibody for Eosinophilic Gastritis and Duodenitis. N Engl J Med 2020;383:1624-1634.

20. (U.S.) NLoM. A Study to Assess AK002 in Eosinophilic Gastritis and/or Eosinophilic Duodenitis (Formerly Referred to as Eosinophilic Gastroenteritis) (ENIGMA 2). [cited 2022 September 5]; Available from: <u>https://clinicaltrials.gov/ct2/show/NCT04322604</u>

21. Talley NJ, Peterson KA, Genta RM, et al. High Discovery Rate of Duodenal and Gastric Eosinophilia in Patients With Unexplained Moderate-Severe Abdominal Symptoms: A Prospective US Multisite Study. Gastroenterology 2023.

22. Dellon ES, Gonsalves N, Rothenberg ME, et al. Determination of Biopsy Yield That Optimally Detects Eosinophilic Gastritis and/or Duodenitis in a Randomized Trial of Lirentelimab. Clin Gastroenterol Hepatol 2022;20:535-545 e15.

23. Fluss R, Faraggi D, Reiser B. Estimation of the Youden Index and its associated cutoff point. Biom J 2005;47:458-72.

24. Lwin T, Melton SD, Genta RM. Eosinophilic gastritis: histopathological characterization and quantification of the normal gastric eosinophil content. Mod Pathol 2011;24:556-63.

25. Lowichik A, Weinberg AG. A quantitative evaluation of mucosal eosinophils in the pediatric gastrointestinal tract. Mod Pathol 1996;9:110-4.

26. Talley NJ, Walker MM, Aro P, et al. Non-ulcer dyspepsia and duodenal eosinophilia: an adult endoscopic population-based case-control study. Clin Gastroenterol Hepatol 2007;5:1175-83.

27. Jenkins D, Goodall A, Gillet FR, et al. Defining duodenitis: quantitative histological study of mucosal responses and their correlations. J Clin Pathol 1985;38:1119-26.

28. DeBrosse CW, Case JW, Putnam PE, et al. Quantity and distribution of eosinophils in the gastrointestinal tract of children. Pediatr Dev Pathol 2006;9:210-8.

29. Hurrell JM, Genta RM, Melton SD. Histopathologic diagnosis of eosinophilic conditions in the gastrointestinal tract. Adv Anat Pathol 2011;18:335-48.

30. Matsushita T, Maruyama R, Ishikawa N, et al. The number and distribution of eosinophils in the adult human gastrointestinal tract: a study and comparison of racial and environmental factors. Am J Surg Pathol 2015;39:521-7.

31. Chernetsova E, Sullivan K, de Nanassy J, et al. Histologic analysis of eosinophils and mast cells of the gastrointestinal tract in healthy Canadian children. Hum Pathol 2016;54:55-63.

32. Lee EH, Yang HR, Lee HS. Quantitative Analysis of Distribution of the Gastrointestinal Tract Eosinophils in Childhood Functional Abdominal Pain Disorders. J Neurogastroenterol Motil 2018;24:614-627.

33. Silva J, Canao P, Espinheira MC, et al. Eosinophils in the gastrointestinal tract: how much is normal? Virchows Arch 2018;473:313-320.

34. Min YW, Lee H, Ahn S, et al. Eosinophil and Mast Cell Counts in the Stomach and Duodenum of Patients with Functional Dyspepsia without a Helicobacter pylori infection. Korean J Gastroenterol 2022;80:28-33.

35. Friesen CA, Sandridge L, Andre L, et al. Mucosal eosinophilia and response to H1/H2 antagonist and cromolyn therapy in pediatric dyspepsia. Clin Pediatr (Phila) 2006;45:143-7.

36. Wauters L, Ceulemans M, Frings D, et al. Proton Pump Inhibitors Reduce Duodenal Eosinophilia, Mast Cells, and Permeability in Patients With Functional Dyspepsia. Gastroenterology 2021;160:1521-1531 e9.

Table 1. Baseline Demographics

Participant characteristics	Pooled symptomatic patients n=707	EoG and/or EoD n=388	Eosinophils <30 n=319	Healthy volunteers n=33
Age, mean (range), y	43 (17-78)	43 (17-78)	42 (18-76)	34 (18-51)
Female sex, n (%)	508 (72)	266 (69)	242 (76)	13 (39)
White, n (%)	626 (89)	336 (87)	290 (91)	33 (100)
Weight, median (range), kg	81 (31-171)	82 (43-171)	79 (31-163)	80 (46-113)
Blood eos ≥250 cells/µL, n (%)	189 (27)	146 (38)	43 (13)	3 (9)
lgE ≥70 kU/L, n (%)	264 (37)	167 (43)	97 (30)	7 (21)
History of atopy, n (%) ^a	357 (50)	225 (58)	132 (41)	5 (15)
History of EoG and/or EoD, n (%) ^b	105 (15)	94 (24)	11 (3)	0 (0)
Gastric eos/5 hpfs, mean \pm SD	25 ± 34	38 ± 41	10 ± 7	7 ± 5
Duodenal eos/3 hpfs, mean ± SD	33 ± 21	45 ± 21	19 ± 7	19 ± 7
TSS [0-60], mean ± SD	29 ± 10	30 ± 10	29 ± 10	0.1 ± 0.2

EoD, eosinophilic duodenitis; EoG, eosinophilic gastroenteritis; eos, eosinophils; hpfs, high-power fields; IgE, immunoglobulin E; kU/L, kilounits per liter; SD, standard deviation; TSS, total symptom score. ^aAsthma, allergic rhinitis, atopic dermatitis, and/or food allergy. ^bA history of the condition was considered to be present if a documented clinical diagnosis had been made by the patient's physician and was noted in the patient's clinical chart history.

FIGURES



Figure 1. Mean tissue eos counts in stomach. Mean of the peak tissue eos counts in 5 hpfs for controls, participants with EoG, and participants with stomach eos counts <30. Caps represent minimum and maximum, boxes represent 25th and 75th percentiles, and the center line corresponds to the median. Welch's unpaired *t* test, *****P*<0.0001. ^a6 outliers in the EoG group not displayed, with mean gastric eos counts of 201, 201, 210, 214, 228, and 300. EoG, eosinophilic gastritis; eos, eosinophils; hpf, high-power field; SD, standard deviation.



Figure 2. Mean tissue eos counts in duodenum. Mean of the peak tissue eos counts in 3 hpfs for controls, participants with EoD, and participants with duodenum eos counts <30. Caps represent minimum and maximum, boxes represent 25th and 75th percentiles, and the center line corresponds to the median. Welch's unpaired *t* test, *****P*<0.0001; ns, not significant. ^a3 patients did not have duodenal biopsy samples (n=2 with EoD; n=1 duodenum <30). ^b2 individuals included in the control group had histologic evidence of EoD (21). EoD, eosinophilic duodenitis; eos, eosinophils; hpf, high-power field; SD, standard deviation.



Figure 3. ROC curves for determining eos count thresholds in 5 gastric hpfs and 3 duodenal hpfs. Optimal eos threshold was determined to be 20 eos/hpf in 5 gastric hpfs for EoG and 33 eos/hpf in 3 duodenal hpfs for EoD. Curves correspond to an AUROC of 0.74 for 3 duodenal hpfs and 0.78 for 5 gastric hpfs. AUROC, area under the receiver operating characteristic; EoD, eosinophilic duodenitis; EoG, eosinophilic gastritis; eos, eosinophil; hpf, high-power field; ROC, receiver operating characteristic.



Figure 4. ROC curves for determining eosinophil count thresholds in 1 gastric hpf and 1 duodenal hpf. Optimal eos threshold was determined to be 33 eos/hpf in 1 gastric hpf for EoG and 37 eos/hpf in 1 duodenal hpf for EoD. Curves correspond to an AUROC of 0.94 for 1 duodenal hpf and 0.94 for 1 gastric hpf. AUROC, area under the receiver operating characteristic; EoD, eosinophilic duodenitis; EoG, eosinophilic gastritis; eos, eosinophil; hpf, high-power field; ROC, receiver operating characteristic.

Determination of Optimal Eosinophil Thresholds for Diagnosis of Eosinophilic Gastritis

and Duodenitis: A Pooled Analysis of 4 Prospective Studies

Supplementary Table. Screening Protocol

Symptom PRO	EGD With Biopsy	Histologic Criteria
 Symptomatic Participants Prior diagnosis or suspected EoG/EoD entered screening TSS6 ≥10 and abdominal pain, diarrhea, or nausea ≥3 on PRO questionnaire 	All Participants Multiple biopsies (≥12) were taken from each participant according to a standardized protocol: • 8-10 gastric biopsies • 4-6 duodenal biopsies • Up to 4 esophageal biopsies (only if participant had a history of EoE or if EoE features were observed during	All Participants Single pathologist evaluated stained biopsy samples and counted eosinophils Entry criteria: • ≥30 eos/hpf in 5 hpfs (stomach) and/or ≥30 eos/hpf in 3 hpfs (duodenum) • No other known cause for GI symptoms or tissue eosinophilia
Healthy Controls	EGD)	

• TSS6 ≤1 EGD, esophagogastroduodenoscopy; EoD, eosinophilic duodenitis; EoE, eosinophilic esophagitis; EoG, eosinophilic gastritis; eos, eosinophils; GI, gastrointestinal; hpf, high-power field; PRO, patient-reported outcome; TSS, total symptom score.

Determination of Optimal Eosinophil Thresholds for Diagnosis of Eosinophilic Gastritis

and Duodenitis: A Pooled Analysis of 4 Prospective Studies



Supplementary Figure. Participant flow chart. ^aThe total study sample reflects individuals with gastric and duodenal biopsy samples. ^bTwo healthy volunteers had histologic evidence of EoD. EoD, eosinophilic duodenitis; EoG, eosinophilic gastritis; TSS6, 6-Point Total Symptom Score.